

Fluorimetric Detection and Quantification of Deoxynivalenol with Zirconyl Nitrate-Ethylenediamine

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Deoxynivalenol (3,7,15-trihydroxy-12,13-epoxytrichothec-9-ene-8-one, vomitoxin) is a cytotoxic trichothecene mycotoxin recently discovered worldwide in grain in association with infestations of Fusarium species, especially in cool, temperate regions (Ueno 1983). Ingestion of this mycotoxin results in feed refusal, emesis, and growth depression in swine and other livestock (Vesonder and Hesseltine 1981), but the specific long-term effects on humans remain unknown.

Quantification of this mycotoxin is relatively difficult because trichothecenes neither fluoresce nor appreciably absorb ultraviolet light (Pathre and Mirocha 1977). Currently, the generally recommended procedure for analysis of deoxynivalenol requires qas-liquid chromatoqraphy with electron-capture detection or gas chromatography - mass spectrometry (Scott 1982). Several colorimetric procedures have been investigated in the search for more practical methods for detection and quantitation and include 4-(p-nitrobenzyl)pyridine, a chromogenic reagent for epoxides (Takitani et al. 1979) and chromotropic acid which reacts with formaldehyde released from trichothecenes upon treatment with sulfuric acid (Kato et al. 1979, Baxter et al. 1983). Type-B trichothecenes such as deoxynivalenol also react with aluminum chloride to produce fluorescent reaction products (Ueno 1973), and this reaction has recently been confirmed for deoxynivalenol (Kamimure et al. 1981). Several type-B trichothecenes, however, produced significantly greater fluorescence upon reaction with zirconyl salts in comparison with aluminum salts and based on this, Kato et al. (1976) developed a fluorimetric assay for fusarenon-X, a toxin related to deoxynivalenol. Through modification of this procedure, we have developed a rapid, simple, sensitive method for the detection and quantitation of deoxynivalenol by fluorimetry.

MATERIALS AND METHODS

Deoxynivalenol (Research Foods Ltd., Downsview, Ontario) standard solution was prepared in methanol at a concentration of 0.04 ug/uL. Fifty microliters (2 ug deoxynivalenol) or less of this standard was added to 2 mL of methanol in 5-mL test tubes. To

this, 25 uL of 5% (v/v) ethylenediamine in methanol and 50 uL of 6% (w/v) zirconyl nitrate in methanol were added. The test tubes were sealed with plastic film, briefly shaken on a vortex mixer, and heated at $40\,^{\circ}\mathrm{C}$ in a water bath for $40\,^{\circ}\mathrm{min}$. After the test tubes were cooled to room temperature, the fluorescence of the reaction product was determined in relation to a reagent blank by using a Turner Model $430\,^{\circ}\mathrm{spectrofluorimeter}$ with excitation at $360\,^{\circ}\mathrm{nm}$ and emission at $454\,^{\circ}\mathrm{nm}$. The half-power band width of the monochrometer was set at $15\,^{\circ}\mathrm{nm}$, and fluorescence intensities were recorded uncorrected.

RESULTS AND DISCUSSION

The fluorescence spectra of the deoxynivalenol reaction product showed excitation maximum at 360 nm and emission maximum at 454 nm. Similar, but not identical values, were reported for related trichothecenes including fusarenon-X, nivalenol, and dehydronivalenol (Kato et al. 1976). The reagent concentrations, however, are critical for production of optimum levels of the fluorescent deoxynivalenol reaction product (Table 1 and 2). Values reported for fusarenon-X (Kato et al. 1976) would produce only 40% of maximum fluorescence for deoxynivalenol.

Table 1. Effect of concentration of ethylenediamine on fluorescence of deoxynivalenol reaction product $^{\rm a}$.

% Ethylenediamine (v/v)	Relative Fluorescence
3.0	39.80
3.5	64.68
4.0	88.06
4.5	97.01
5.0	100.00
5.5	99.50
6 . D	92.04
6.5	89.85

 $^{^{}a}$ 25 uL deoxynivalenol (1 ug); 25 uL ethylenediamine; 50 uL 6% (w/v) zirconyl nitrate; 2 mL methanol; 40 $^{\circ}$ C for 40 min. Values are means of three determinations.

Table 2. Effect of concentration of zirconyl nitrate on fluorescence intensity of deoxynivalenol reaction product^a.

% Zirconyl Nitrate (w/v)	Relative Fluorescence
3.0	57.46
3.5	65,61
4.0	69.40
4.5	75.11
5.0	80.90
5.5	88.51
6.0	100.00
6.5	99.57
7.0	95.48
8.0	81.90
9.0	65.15

 a 25 uL deoxynivalenol (1 ug); 25 uL of 5% (v/v) ethylenediamine; 50 uL zirconyl nitrate; 2 mL methanol; 40 $^{\circ}$ C for 40 min. Values are means of three determinations.

We chose a 40 min reaction time at 40°C . Shorter durations are required at higher temperatures but the risk of thermal decomposition of the reaction product is also apparent(Fig. 1). Reactions at higher temperatures might be of interest for possible application to post-column derivitization to improve minimum detectability of deoxynivalenol following high performance liquid chromatography (HPLC).

The fluorescent reaction product was stable at room temperature for at least 4 h. Calibration curves showed excellent linearity from 25-2000 ng/2 mL methanol (r=0.996). Individual determinations were highly reproducible with a coefficient of variation of 1.33% and 2.71% for 1000 and 100 ng, respectively (calculated for 10 determinations).

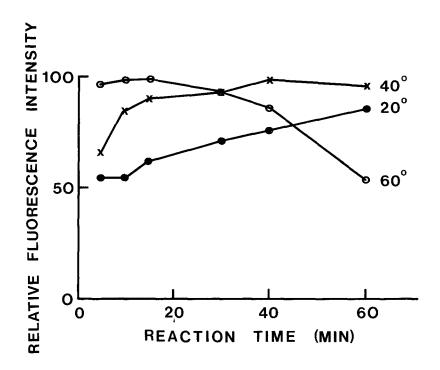


Figure. 1 Effect of incubation temperature and duration of heating on fluorescence intensity of deoxynivalenol reaction product.

This determination method for deoxynivalenol is rapid, sensitive, accurate and economical to perform. In view of its efficiency and practicality, further work is warranted on its possible adaptation as a determinative procedure in analytical methods to screen agricultural commodities.

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REFERENCES

Baxter JA, Terhune SJ, Qureshi, SA (1983) Use of chromotropic acid for improved thin-layer chromatographic visualization of trichothecene mycotoxins. J Chromatogr 261:130-133

Kamimura H, Nishijima M, Yasuda K, Saito K, Ibe A, Nagayama T, Ushiyama H, Naoi Y (1981) Simultaneous detection of several Fusarium mycotoxins in cereals, grain, and foodstuffs. J Assoc Off Anal Chem 64:1067-1073

- Kato T, Asabe Y, Suzuki M, Takitani S (1976) Fluorimetric determination of fusarenon-X and its related mycotoxins. Bunseki Kagaku 25:659-662
- Kato T, Asabe Y, Suzuki M, Takitani, S (1979) Spectrophotometric and fluorimetric determinations of trichothecene mycotoxins with reagents for formaldehyde. Anal Chim Acta 106:59-65
- Pathre SV, Mirocha CJ (1977) Assay methods for trichothecenes and review of their natural occurrence. In: Rodricks JV, Hesseltine CW, Mehlman MA (eds) Mycotoxins in human and animal health. Pathotox Publishers, Park Forest South, IL p 229
- Scott PM (1982) Assessment of quantitative methods for determination of trichothecenes in grain and grain products. J Assoc Off Anal Chem 65:876-883
- Takitani S, Asabe Y, Kato T, Suzuki M, Ueno Y (1979) Spectrodensitometric determination of trichothecene mycotoxins with 4-(p-nitrobenzyl)pyridine on silica gel thin-layer chromatograms. J Chromatogr 172:335-342
- Ueno Y (1983) Trichothecenes chemical, biological and toxicological aspects. Dev Food Sci 4:1-313
- Ueno Y, Sato N, Ishii K, Sakai K, Tsunoda H, Enomoto M (1973) Biological and chemical detection of trichothecene mycotoxins of Fusarium species. Appl Microbiol 25:699-704
- Vesonder RF, Hesseltine CW (1981) Vomitoxin: natural occurrence on cereal grain and significance as a refusal and emetic factor to swine. Process Biochem 16:13-15

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